Protocol for DNA purification from formalin-fixed paraffin-embedded tissue

1. Cut a small section (10 - 20 mg) of the formalin-fixed paraffin-embedded tissue and manually remove any excess of paraffin as much as possible. Place the tissue in a 2 ml tube.

2. Add 1.2 ml Xylene to the sample and vortex vigorously.

3. Centrifuge at full speed for 5 min at room temperature.

4. Carefully remove the supernatant by pipetting. Do not remove the pellet.

5. Add 1.2 ml ethanol (96 - 100 %) to the pellet, and mix gently by vortexing.

6. Centrifuge at full speed for 5 min at room temperature.

7. Carefully remove the ethanol by pipetting. Do not remove the pellet. Repeat steps 5-7 once.

8. Incubate the tube containing the sample (with the cap open) at 37°C for 10 minutes until the ethanol has evaporated.

9. Immediately resuspend the tissue pellet in 50 - 200 µl TE Buffer. Homogenize and process the tissue according to the protocol described in the QuickPick™ SML gDNA kit insert.

Table 1. Reagent volumes for genomic DNA purifications

Reagent	**Reagent volume per preparation**			
Sample amount	**1.25 mg**	**2.5 mg**	**5 mg**	**10 mg**
Lysis Buffer	25 µl	50 µl	100 µl	200 µl
Proteinase K	2.5 µl	5 µl	10 µl	20 µl
Binding Buffer	62.5 µl	125 µl	250 µl	500 µl
Magnetic Particles	2 µl	4 µl	8 µl	16 µl
Wash Buffer 1	2 x 125 µl	2 x 250 µl	2 x 500 µl	2 x 750 µl
Wash Buffer 2	125 µl	250 µl	500 µl	750 µl
Elution Buffer	5 - 25 µl	10 - 50 µl	25 - 100 µl	50 - 200 µl

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